

DOI: 10.14744/SEMB.2020.18794 Med Bull Sisli Etfal Hosp 2020;54(2):159–168

## Review



# Pheochromocytoma and Paraganglioma: From Epidemiology to Clinical Findings

### 💿 Nurcihan Aygun, 💿 Mehmet Uludag

Department of Genaral Surgery, Health Sciences University, Sisli Hamidiye Etfal Medical Practice and Research Center, Istanbul, Turkey

#### Abstract

Pheochromocytomas (PCC) and paragangliomas (PGL) are rare neuroendocrine tumors. Pheochromocytomas arise from chromaffin cells in the adrenal medulla, and PGLs arise from chromaffin cells in the ganglia of the autonomic nervous system. Paragangliomas originate from sympathetic or parasympathetic ganglia in the abdomen, thorax, and pelvis. The majority of PCC and sympathetic PGL are endocrine active tumors causing clinical symptoms by secreting excess catecholamines (norepinephrine, epinephrine, dopamine) and their metabolites. The incidence of PCC and PGL ranges between 2 and 8 per million, with a prevalence between 1:2500 and 1:6500. It peaks between the 3rd and 5th decades of life, and approximately 20% of cases are pediatric patients. The prevalence among patients with hypertension in outpatient clinic ranges between 0.1-0.6% in adults and between 2-4.5% in the pediatric age group. 10-49% of these tumors is detected incidentally in imaging techniques performed for other reasons. However, 4-8% of adrenal incidentalomas are PCCs. Of these neuroendocrine tumors, 80-85% are PCCs and 15-20% are PGLs. Up to 40% of patients with PCC and PGL has disease-specific germline mutations and the situation is hereditary. Of 60% of the remaining sporadic patients, at least 1/3 has a somatic mutation in predisposing genes. 8% of the sporadic cases, 20-75% of the hereditary cases, 5% of the bilateral, adrenal cases, and 33% of the extra-adrenal cases at first presentation are metastatic. Although PCCs and PGLs have scoring systems for histological evaluation of the primary tumor, it is not possible to diagnose whether the tumor is malignant since there is no histological system approved for the biological aggressiveness of this tumor group. Metastasis is defined as the presence of chromaffin tissue in non-chromaffin organs, such as lymph nodes, liver, lungs and bone. Although most of the PCC and PGL are benign, the metastatic disease may develop in 15-17%. Metastatic disease is reported between 2-25% in PCCs and 2.4-60% in PGLs. The TNM staging system of the American Joint Committee on Cancer (AJCC) was developed to predict the prognosis, based on the specific anatomical features of the primary tumor and the occurrence of metastasis. Keywords: Catecholamine synthesis and metabolism; paraganglioma; pheochromocytoma.

Please cite this article as "Aygun N, Uludag M. Pheochromocytoma and Paraganglioma: From Epidemiology to Clinical Findings. Med Bull Sisli Etfal Hosp 2020;54(2):159–168".

Pheochromocytoma (PCC) and paraganglioma (PGL) are rare neuroendocrine tumors. Pheochromocytomas arise from chromaffin cells in the adrenal medulla, and PGLs arise from chromaffin cells in the ganglia of the autonomic nervous system.<sup>[1, 2]</sup> Paragangliomas originate from sympathetic or parasympathetic ganglia in the abdomen, thorax, and pelvis. Paragangliomas may also originate from parasympathetic ganglia at the base of the skull and along the glossopharyngeal nerve and vagus nerve in the neck.<sup>[2]</sup> The majority of PCCs and sympathetic PGLs are endocrine active tumors, and they cause clinical symptoms by secreting excess catecholamines (norepinephrine, epinephrine,

Address for correspondence: Nurcihan Aygun, MD. Sisli Hamidiye Etfal Tip Uygulama ve Arastirma Merkezi, Saglik Bilimleri Universitesi, Genel Cerrahi Anabilim Dali, Istanbul, Turkey

Phone: +90 553 277 95 78 E-mail: nurcihanaygun@hotmail.com

Submitted Date: May 16, 2020 Accepted Date: May 27, 2020 Available Online Date: June 03, 2020 ©Copyright 2020 by The Medical Bulletin of Sisli Etfal Hospital - Available online at www.sislietfaltip.org OPEN ACCESS This is an open access article under the CC BY-NC license (http://creativecommons.org/licenses/by-nc/4.0/).



dopamine) and their metabolites. However, approximately 2/3 of PGLs of parasympathetic origin in the head and neck region are not endocrine active.<sup>[3]</sup> Paragangliomas are sometimes called extra-adrenal PCCs. These tumors are abbreviated in the literature as PCC/PGL or PPGL; in this study, PPGL abbreviation will be used.<sup>[1, 2, 4]</sup>

## **Epidemiology and Incidence**

The incidence of PPGL ranges between 2 and 8 per million, with a prevalence between 1:2500 and 1:6500.<sup>[5, 6]</sup> The annual incidence in the USA is approximately 500-1600 cases per year, which is lower than the prevalence of PPGL (0.05-0.1%) detected in the autopsy series incidentally.<sup>[6]</sup> PPGL peaks between the 3rd and 5th decades of life, and approximately 20% of cases is pediatric patients.<sup>[5, 7]</sup> The prevalence among patients with hypertension in outpatient clinic ranges between 0.1-0.6% in adults and between 2-4.5% in the pediatric age group.<sup>[8]</sup> 10-49% of PPGLs is detected incidentally in imaging techniques performed for other reasons. However, 4-8% of adrenal incidentalomas are PCCs.<sup>[5]</sup> Of these neuroendocrine tumors, 80-85% is PCCs, and 15-20% is PGLs.<sup>[1, 2]</sup> 70-80% of PGLs arises from infradiaphragmatic sympathetic ganglia. It most commonly originates from the Zuckerkandl organ located near the origin of the inferior mesenteric artery at the aortic bifurcation. It can be less frequently found in the retroperitoneal area around the kidney and adrenal and in the bladder; however, it can be seen in all anatomical areas in the abdomen where sympathetic or parasympathetic ganglia are localized. Approximately 10% of the PGLs are in the mediastinal and pericardial localization within the thorax, and approximately 20% is localized in the head and neck region.<sup>[2, 9]</sup> Up to 40% of patients with PPGL have diseasespecific germline mutations and the situation is hereditary. Of the 60% of the remaining sporadic patients, at least 1/3 has a somatic mutation in predisposing genes.<sup>[10]</sup> 8% of the sporadic cases, 20-75% of the hereditary ones, 5% of the bilateral, adrenal cases, and 33% of the extradrenal ones at first presentation are metastatic.<sup>[5]</sup>

Although the 10's rule, which Graham defined in 1951 for PPGL, describing 10% extradrenal, 10% bilateral, and 10% malignancy, is still included in many textbooks, it has expired today.<sup>[11]</sup>

#### **Catecholamine Synthesis and Metabolism**

Synthesis of catecholamine hormones takes place in the adrenal medulla, sympathetic and central nervous system. Catecholamine synthesis begins with the conversion of L-tyrosine, which is taken from the diet or synthesized from phenylalanine in the liver, to L- dihydroxyphenylalanine (DOPA) by catalyzation of the tyrosine hydroxylase enzyme.

<sup>[1]</sup> This stage is the rate-limiting step of catecholamine synthesis; hence, the source of the catecholamines depends on the presence of this enzyme.<sup>[12]</sup> Tyrosine hydroxylase enzyme is largely found in the catecholaminergic nerve endings of the brain and in the sympathetic nervous system in the periphery, in the adrenal medulla, and extraadrenal chromaffin cells.<sup>[12]</sup> DOPA is converted into dopamine with L-aromatic amino acid decarboxylase enzyme. Dopamine is transported by vesicular monoamine transporter into storage vesicles inside the cell. Dopamine is converted intravesically to norepinephrine with a dopamine-betahydroxylase enzyme, which is one of the components of storage vesicles (Fig. 1).<sup>[1, 12]</sup> Norepinephrine is converted to epinephrine (adrenaline) with the phenylethanolamine N-methyl transferase (PNMT) enzyme found only in the adrenal medulla and the Zuckerkandl organ, sympathetic ganglion (Fig. 1). Since PNMT is a cytosolic enzyme, epinephrine synthesis is dependent on the infiltration of norepinephrine from its synthesis site, vesicles to the cytoplasm. There are two subgroups of chromaffin cells. Adrenergic cells (epinephrine cells) contain PNMT and store mainly epinephrine, and these cells contain medium-density large granules containing epinephrine.<sup>[12]</sup> Adrenergic cells are localized close to the cortex and are exposed to high cortisol levels of the cortex. Cortisol is required for the induction of the PNMT enzyme.<sup>[13]</sup> Norepinephrine cells do not contain PNMT; they mainly store norepinephrine and contain small electron density granules containing norepinephrine.<sup>[12]</sup> Failure to secrete epinephrine in extra-adrenal tissues may also arise from the absence of cortisol for PNMT

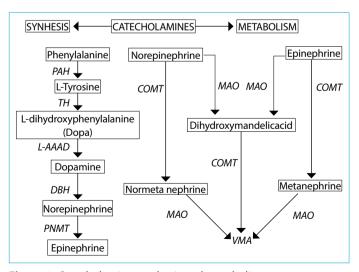


Figure 1. Catecholamine synthesis and metabolism.

PAH: Phenylalaninehydroxylase; TH: Tyrosine hydroxylase; L-AAAD: L- aromatic amino acid decarboxylase; DBH: Dopamine-beta-hydroxylase; PNMT: Phenyletanolamine-N-methyltransferase; MAO: Monoamine oxidase; COMT: Catechol O-methyltransferase; VMA: (vanillylmandelic acid) 3-methoxy-4-hydroxymandelic acid. stimulation.<sup>[13]</sup> The circulating epinephrine is secreted by the adrenal medulla, is released directly from the medulla to the circulation, creating a hormonal effect on cells in other tissues and its basal value is 0.1-0.3 nmol/L. Norepinephrine acts as a neurotransmitter in some neurons in the brain and peripheral sympathetic neurons. Norepinephrine is secreted into the synaptic cleft for activation and taken back into the nerve from the synaptic cleft. As a result, a small amount of norepinephrine is released into the circulation by escaping from the sympathetic nerves. More than 90% of circulating norepinephrine originates from sympathetic nerves and approximately 7% from the adrenal medulla and innervates the mesenteric organs, kidney and skeletal muscles. The plasma level in basal conditions is 0.6-2 nmol/L. However, circulating epinephrine is released only from the adrenal medulla, and its level is 0.1-0.3 nmol/L. Dopamine, which is not converted to norepinephrine and epinephrine, is metabolized in the gastrointestinal tract.<sup>[12,</sup> <sup>13]</sup> When the adrenal medulla is stimulated, the secretory vesicles join the cell membrane in chromaffin cells, and the entire vesicle content is released to the circulation through its rich vascular network. These vesicles contain catecholamines, as well as ATP, calcium, magnesium, chromogranins and other neuropeptides.<sup>[14]</sup> Catecholamine content may vary depending on enzyme expression in PCCs. Most PCCs produce predominantly norepinephrine. Some produce both norepinephrine and epinephrine, and rarely epinephrine. Since dopamine is predominantly converted to norepinephrine, in very rare cases, the mainly produced molecule is dopamine.<sup>[12]</sup> Since there is no PNMT in PGLs, norepinephrine is produced predominantly.

Catecholamines show their biological effects by binding to transmembrane adrenergic receptors, which are G proteincoupled receptors. There are adrenergic receptors with different affinity for catecholamines in different tissues and these are  $\alpha$ ,  $\beta$ 1,  $\beta$ 2 receptors. The receptor affinities for  $\alpha$  receptors are; epinephrine > norepinephrine >> isoproterenol; for  $\beta$ 1 receptors; isoproterenol > epinephrine = norepinephrine; and for  $\beta$ 2 receptors; isoproterenol > epinephrine >> norepinephrine.<sup>[14]</sup> The effects of catecholamines in the organs of where receptors are present through these receptors can be summarized as follows: While a1 receptors cause vasoconstriction in the blood vessels, decreased motility in the intestine, increased sphincter tone, decreased insulin and glucagon release in the pancreas, glycogenolysis and gluconeogenesis in the liver, dilation of the pupils, uterine contraction and sweating, α2 receptors inhibit norepinephrine release in synapses and stimulate platelet aggregation. While B1 receptors cause chronotropic and inotropic effects on the heart, vasodilation in coronary vessels, lipolysis in adipose tissue, decreased motility and increased

sphincter tone in the intestine, increase in insulin and glucagon release in the pancreas;  $\beta 2$  receptors cause vasodilation in the blood vessels, bronchodilation in the lungs and uterine relaxation.<sup>[14]</sup>

Many of the catecholamines are metabolized in the neural cells in which they are produced; catecholamines produced in the adrenal medulla are metabolized with adrenal medulla. The stimulation half-life of the hormones released to the blood is very short, about 1-2 minutes, and they are taken by the non-neural tissues. The metabolism of catecholamines given to circulation takes place primarily in the liver and kidneys. Norepinephrine and epinephrine are metabolized to normetanephrine and metanephrine with catechol-o-methyltransferase (COMT) in the adrenal medulla and non-neural tissues. These metabolites are metabolized to the end product, vanillylmandelic acid (VMA), with monoamine oxidase enzyme (MAO). Norepinephrine and epinephrine can also be metabolised to VMA using MAO and then with COMT. VMA is excreted in urine as conjugated with sulfuric and glucuronic acid, which happens in the liver and intestine. Small amounts of free dopamine, epinephrine and norepinephrine are excreted in the urine (Fig. 1).<sup>[1, 12, 13]</sup> Catecholamines are secreted from adrenal medulla intermittently and their half-life is very short. Therefore, these may reflect the sympathoadrenal activity at the time the sample was taken. Oxymethyl derivatives of catecholamines are continuously produced in tumor cells. The plasma half-life of these metabolites is much longer than norepinephrine and epinephrine. Their plasma levels are relatively more stable. Plasma metanephrine level is often independent of sympathoadrenal stimulation.<sup>[1, 13]</sup>

### Pathophysiology

PPGLs release an uncontrolled excess of catecholamines, which bind to receptors in different organs, causing some physiological changes. Norepinephrine stimulates  $\alpha 1$ ,  $\alpha 2$ , and  $\beta$ 1 receptors, whereas epinephrine stimulates only  $\beta$ 1 and B2 receptors. While normal levels of dopamine do not affect adrenergic receptors, it can stimulate both  $\alpha$  and  $\beta$  receptors when dopamine increases in circulation, as in case of tumors that secrete dopamine. The a1 adrenergic receptor is primarily located in the smooth muscles of the peripheral arteries and veins (such as coronary, peripheral and renal), and their stimulation causes vasoconstriction. Presynaptic a2 adrenergic receptors inhibit norepinephrine secretion in sympathetic ganglia. Stimulation of a2 receptors in smooth muscles causes arterial vasodilation and coronary vasoconstriction.  $\beta$ 1 receptors are primarily localized in the heart and have positive inotropic and chronotropic effects on the heart. In addition, B1 receptors increase renin secretion, resulting in an increase in mean arterial pressure. β2 receptors

Patients with PPGL with these mutations can be divided

lead to vasodilation in skeletal muscle arteries. It increases the release of norepinephrine from sympathetic nerve endings. Dopamine stimulates D1 and D2 receptors. D1 receptors cause vasodilation in the renal arteries. Stimulation of D2 receptors inhibits the secretion of norepinephrine in the sympathetic nerve endings, and it has a slightly negative inotropic effect on the heart. In addition, high levels of dopamine stimulate  $\alpha$  and  $\beta$ 1 receptors, causing vasoconstriction and increased heart rate.<sup>[15, 16]</sup>

PCCs often secrete both epinephrine and norepinephrine at different rates. PGLs, on the other hand, are not able to synthesize epinephrine since they do not contain the PNMT enzyme. They only secrete norepinephrine. Different catecholamines and their ratios may cause different vasoactive effects. In some cases, clinical findings can be extremely informative.

Prolonged and recurrent norepinephrine secretion results in prolonged vasoconstriction and increased blood pressure, and venous vasoconstriction results in decreased blood volume in the circulation. If preoperative a blockade is not applied and volume support is not provided in these patients, acute hypovolemia may occur as the vasoconstriction caused by norepinephrine disappears after PPGL extraction in surgery. In tumors that predominantly secrete epinephrine, tachyarrhythmia arises in addition to hypertension.<sup>[1, 5]</sup> In addition, postural hypotension occurs due to vasodilation in skeletal muscles with epinephrine stimulation. Hypertension and palpitations may not be present in tumors producing dopamine.<sup>[2, 15]</sup> Increased plasma catecholamine level, increased glycogenolysis and inhibition of insulin secretion in pancreatic islet cells result in findings of diabetes. Increased catecholamines can cause stress-induced cardiomyopathy (Takotsubo cardiomyopathy).<sup>[5]</sup>

The name pheochromocytoma crisis is the name given to a group of symptoms. It is a rare condition resulting from uncontrolled catecholamine secretion characterized by multisystem organ failure, high fever, encephalopathy, severe hypertension or hypotension. If not recognized and not treated urgently, it may progress to metabolic acidosis and lead to death.<sup>[2, 5]</sup>

## **Genetic Basis and Genetic Testing**

Up to 40% of patients with PPGL have disease-specific germline mutations and are one of the most common hereditary tumors. At least 1/3 of the remaining 60% sporadic cases have a somatic mutation in predisposing genes and approximately 60% of patients with PPGL have germline or somatic mutation. More than 20 gene mutations have been detected to date.<sup>[10, 17, 18]</sup>

into three groups according to the cancer genome map. These groups are:  $\ensuremath{^{[19]}}$ 

Cluster 1 - pseudohypoxic Krebs cycle-related genes

Cluster 2 - Kinase Signaling-Related Genes

Cluster 3 - Wnt Signaling-Related Genes

### **Cluster 1 - Pseudohypoxic Krebs Cycle-Related Genes**

This group is called the pseudohypoxic pathway because it mimics cellular hypoxia.<sup>[20]</sup>

This group can be divided into two main subgroups. Those associated with tricarboxylic acid cycle contain germline mutations in genes encoding SDHx (succinate dehydrogenase) subunits (SDHA, SDHB, SDHC, SDHD), SDHAF2 (succinate dehydrogenase complex assembly factor-2), FH (fumarate hydratase), MDH2 (malate dehydrogenase 2), IDH (isocitrate dehydrogenase 1) and the majority of patients with hereditary PGL are associated with these mutations. Due to mutations in genes encoding enzymes in this group, the phosphorylation in the Krebs cycle is disrupted, glycolysis increases and results in the accumulation of oncometabolites, such as succinate, malate and fumarate. PPGLs, especially related to mutations in SDHx subunits, are often multiple, aggressive and metastatic tumors compared to those originating from other mutations, especially cluster 2 mutations.<sup>[7]</sup>

Mutations related to the other subgroup VHL/EPAS1 genes contain germline mutations in PHD1/2 (EgI-9 prolyl hydroxylase-1 and -2), VHL (von Hippel–Lindau tumor suppressor) and HIF2A/EPAS1/2 (hypoxia-inducible factor 2 $\alpha$ ) genes. 95-100% of patients with VHL syndrome are related to the mutation in the VHL tumor suppressor gene. By reduction in hydroxylation and reduction of these HIF proteins, HIF- $\alpha$ stabilizes and accumulates.<sup>[20]</sup>

This results in cellular proliferation, decreased apoptosis and angiogenesis. EPAS1 related gene mutations also increase the transcription of HIF-2 $\alpha$  genes, similar to VHL.<sup>[7]</sup> In addition, it has recently been suggested that mutations in EPAS genes have unique molecular signatures in PPGLs.<sup>[21]</sup>

There were different effects of genes related to HIF transcription, oxidative phosphorylation, and angiogenesis compared with VHL- and SDHx-related PPGLs. In the future, mutations in VHL and EPAS genes can be divided into two specific subgroups.<sup>[7, 20]</sup>

Almost all tumors with mutations in cluster 1 are tumors with a noradrenergic biochemical phenotype. These produce norepinephrine and dopamine, not epinephrine.<sup>[20]</sup>

## **Cluster 2 - Kinase Signaling-Related Genes**

Cluster 2 kinase signaling pathway is associated with germ-

line or somatic mutations in the RET (rearranged-duringtransfection) proto-oncogene, NF 1 (neurofibromin 1) tumor suppressor, HRAS, TMEM127 (transmembrane protein 127) and MAX (Myc-associated factor X) genes. Mutations in these genes stimulate cell proliferation, cell surveillance, cancer development, and angiogenesis by activating PI3K/AKT (phosphatidylinositol-3-kinase), RAS/RAF/ERK, mTORC1 (mammalian target of rapamycin)/p70S6K (p70S6 kinase) signaling pathways.<sup>[20]</sup>

The most common hereditary syndrome in this group is MEN syndromes.<sup>[20]</sup> Many of the cluster 2 mutations have adrenergic biochemical phenotype and are associated with increased epinephrine production as a result of high PNMT expression. Norepinephrine may increase or noradrenaline may be at normal levels.<sup>[20]</sup>

Most PCCs associated with these mutations are benign but have a high rate of recurrence and multifocality.<sup>[22]</sup>

### **Cluster 3 - Wnt Signaling-Related Genes**

CSDE1 (Cold Shock Domain Containing E1) gene and the MAML3 (mastermind-like transcriptional coactivator 3) fusion gene contain PCCs with somatic mutations that activate the Wnt and Hedgehog (Hh) signaling pathways. These tumors strongly express chromogranin A.<sup>[20]</sup> It has been suggested that this category is related to aggressive PPGLs, because some of the patients with this mutation admitted with recurrence and metastasis.<sup>[19]</sup>

The germline mutation of this category is unknown.<sup>[22]</sup> The biochemical phenotype of this group is also unknown.<sup>[20]</sup>

In general, the risk of metastatic disease from high to low is as follows: mutations associated with cluster 1 tricarboxylic acid cycle, mutations associated with group 1VHL/ EPAS1 genes, cluster 3, cluster 2.<sup>[7]</sup>

## Genetic Syndromes Associated with Pheochromocytoma and Paraganglioma

#### Multiple Endocrine Neoplasia Type 2 (MEN 2) (Table 1)

MEN2 syndromes are autosomal dominant diseases associated with RET proto-oncogene mutation located on chromosome 10. There is a strong phenotype-genotype relationship in these syndromes. More than a hundred RET point mutations have been identified, approximately 90% have single point mutations.<sup>[23]</sup>

There are three subtypes of MEN syndrome. 75% of MEN syndromes are MEN2A, 5% are MEN2B and 20% are familial medullary thyroid cancer. The familial medullary thyroid cancer subgroup has no known components other than medullary cancer.<sup>[24]</sup>

Medullary cancer develops in almost 100% of the patients with MEN2A, PCC in 50%, primary hyperparathyroidism in 10%. Approximately 100% of the patients with MEN2B develop medullary cancer, 50% develop PCC, and 95% develop the gastrointestinal system, mucosal neuroma in the mouth and lips and Marfanoid appearance.<sup>[24]</sup> In MEN syndromes, the tumor is often localized in the adrenal medulla and PGL is very rare.<sup>[7]</sup> PCCs in this syndrome make up 5% of all PCCs.<sup>[5]</sup> Bilateral adrenal involvement occurs in approximately 50-60%.<sup>[25, 26]</sup> Bilateralism can be synchronous or metachronous.<sup>[27]</sup> Its biochemical phenotype is adrenergic.<sup>[28]</sup>

Metastatic pheochromocytoma is seen below 5%.<sup>[5, 25]</sup> In addition, very rarely, head and neck PGL is defined in MEN2 syndromes.<sup>[28]</sup>

PCC should be investigated before any intervention in MEN2 syndrome, and if any, PCC should be treated before surgery for other components of the syndrome.

#### Von Hippel Lindau Syndrome (Table 1)

Von Hippel Lindau Syndrome is an autosomal dominant syndrome that arises from the germline mutation in the VHL tumor suppressor gene on the 3p25-26 chromosome. <sup>[25]</sup> Several different mutations in the VHL gene may cause this syndrome and affect multiple organs. Its prevalence is one in 36000 people.<sup>[24]</sup> Cerebellar and spinal hemangioblastoma, retinal hemangioma, renal cysts, clear cell renal carcinoma, pancreatic neuroendocrine tumors, cysts and cystadenomas of the pancreas, PPGLs, cystadenomas in gonads, benign asymptomatic lung lesions, benign asymptomatic liver lesions, liver cysts, endolymphatic cystic tumors can be seen in this syndrome.<sup>[25]</sup> In 20% of the patients with VHL, PCC usually occurs and is seen at a young age. Half of the pediatric age group, which presents with PPGL, has VHL syndrome.<sup>[2]</sup>

Patients with PPGL with VHL syndrome constitute 5-10% of all PPGL patients.<sup>[5]</sup> 40-50% of PCCs in this syndrome are bilateral.<sup>[25]</sup> The biochemical phenotype is noradrenergic. Metastatic PCC is rare.<sup>[2]</sup> In VHL syndrome, PGL is rare, usually of parasympathetic origin and does not secrete a significant amount of catecholamines.<sup>[29]</sup> VHL syndrome is divided into two subgroups, as type 1 if there is no PCC or type 2 if PCC is present. Type 1 includes cerebellar and spinal hemangioblastoma, retinal hemangioma, multiple abdominal cysts and tumors, clear cell renal carcinoma, but no PCC. Type 2 is divided into three types according to the presence and aggressiveness of renal carcinoma. Cerebellar and spinal hemangioblastoma, retinal hemangioma, low-risk renal carcinoma and PCC are present in Type 2A, cerebellar and spinal hemangioblastoma, retinal hemangioma, retinal hemangioma, low-risk renal carcinoma and PCC are present in Type 2A, cerebellar and spinal hemangioblastoma, retinal hemangioblastoma, retinal hemangioma, retinal hemangioma, low-risk renal carcinoma and PCC are present in Type 2A, cerebellar and spinal hemangioblastoma, retinal hemangioblastoma, retinal hemangioblastoma, retinal hemangioma, low-risk renal carcinoma and PCC are present in Type 2A, cerebellar and spinal hemangioblastoma, retinal hemangioblastoma, retinal hemangioblastoma, retinal hemangioblastoma, retinal hemangioblastoma, retinal hemangioma, low-risk renal carcinoma and PCC are present in Type 2A, cerebellar and spinal hemangioblastoma, retinal he

Syndrome	Subtype	Mutation Gen	Biochemical Phenotype	PCC/PGL penetrance	Penetrance PCC	Penetrance pPGL	Penetrance sPGL	Frequency All PCC	Bilaterality	Frequency Bilaterality Proportion Other All PCC Malignant Manif	Proportion Other Malignant Manifestations
MEN	MEN2A MEN2B	RET (10q11.2)	A	50%	High	Very low	Very low	%5	50-60%	<5%	MTC pHPT MTC
											Mucosal neuroma, colonic ganglioneuromatosis, marfanoid facies,
VHL	Type 1	VHL (3p25-p26)	NA	20%	High	Very low	Low	5-10%	40-50%	<5%	Hemangioblastoma in the retina and central
											nervous system, multiple abdominal neoplasms and cvsts. RCC and PCC
NF1		NF1 (17a11.2)	<	1-5%	Low	Verv low	Verv low	1%	20%	10%	Cafe-au-lait macules.
											malign glioma, neurofibromas, freckling
											in the axillary or inguinal
											region, GIST, osseous lesion, and PCC
FPGL	FPGL1	SDHD (11q23)	NA, D	unknown	Low	High multifocal	Low	5-10%	low	<5%	RCC
											GIST
	FPGL2 S	SDHAF2 (11q13.1)	) NA, A	unknown	Very low	High multifocal	Very low	rare		unknown	pituitary adenoma
	FPGL3	SDHC (1q21)	NA, D	unknown	Low	Low	Low	rare		unknown	
	FPGL4	SDHB (1p35-p36)	NA, D	unknown	Low	Intermediate Intermediate	Intermediate	10-20%	low	30-70%	
	FPGL5	SDHA (5p15.33)	NA, D	unknown	Very low	Very low	Very low	<5%	low	10%	

gioma, high-risk renal carcinoma, endocrine neoplasms, pancreatic cysts and PCC are present in Type 2B and only PCC is present in Type 2C.<sup>[29]</sup>

Cases with PCC are often asymptomatic and, unlike sporadic PCCs, they do not release excess catecholamines.<sup>[29]</sup>

## Neurofibromatosis 1 (Von Recklinghausen's Disease (VRD) (Table 1)

Neurofibromatosis 1 is an autosomal dominant syndrome that arises from a mutation in the NF1 gene, and its prevalence is 1 in 3000 people. This gene is localized on the 17qll.2 chromosome and encodes the neurofibromin protein. In the absence or decreased levels of neurofibromin with mutations in the NF1 gene, cell proliferation through various pathways and inhibited apoptosis occur.<sup>[30]</sup> NF1 is characterized by multiple neuromas and is characterized by 15% peripheral nerve sheath tumors.<sup>[2]</sup> In addition, caféau-lait spots, freckles in the axilla and inguinal areas, malignant glioma, bone lesions, gastrointestinal stromal tumors, and PPGL may occur.<sup>[5, 31]</sup>

PPGL develops in 1-5% of patients with NF-1. Almost all of these tumors are PCC, and PGL occurs rarely. They constitute 1% of all patients with PCC.<sup>[5]</sup> They are 20% bilateral.<sup>[1]</sup>

Approximately 10% of them are malignant.<sup>[2, 5]</sup>

## Hereditary Paraganglioma Syndromes (PGL Syndromes) TYPE 1-5 (SDH COMPLEX) (Table 1)

There are four subunits of succinate dehydrogenase (SDHA, SDHB, SDHC, SDHD), which cause syndromes with the autosomal dominant inheritance that occur with germline mutations in genes on different chromosomes that encode different subunits. Type 1 is associated with SDHD mutations, type 2 with SDHAF2 mutations, type 3 with SDHC mutations, type 4 with SDHB mutations, and type 5 with SDHA mutations.<sup>[5, 31]</sup> The prevalence is estimated as 1 in 1 million people.<sup>[24]</sup> These syndromes are more often associated with PGL. Head and neck PGLs are common. PCCs are seen in all five types. Malignancy rate is higher in SDHB mutations and 30-70% malignancy has been reported. In other types, the malignancy rate is low.<sup>[2]</sup>

Apart from these, renal cell carcinoma, gastrointestinal stromal tumors, pituitary adenomas can be seen. There is a risk of papillary thyroid cancer and breast cancer in SDHB mutations.<sup>[32]</sup>

## Other Mutations Associated with Pheochromocytoma/ Paraganglioma

PPGL can also be seen in TMEM127 mutations, MAX mutations, KIF1B mutations, EGLN1 mutations, MDH2 mutations, FH mutations, KIF1 $\beta$  mutations, HIF2A, CSDE1, MAML3 Pacak-Zhuang Syndrome, Carney complex, Carney-Stratakis Syndrome, MEN1 Syndrome.<sup>[1, 20, 31]</sup>

## Clinical and Biological Behavior of Pheochromocytoma/ Paraganglioma

Although PPGLs have scoring systems for histological evaluation of the primary tumor, it is not possible to diagnose whether the tumor is malignant, since there is no histological system approved for the biological aggressiveness of this tumor group.[33] Malignancy in these tumors can be defined as metastasis at presentation or during subsequent follow-up. Metastasis is defined as the presence of chromaffin tissue in non-chromaffin organs, such as lymph nodes, liver, lungs and bone.[34] The proposed benign and malignant definitions for PCC in the 3<sup>rd</sup> version of the World Health Organization (WHO) published on the classification of endocrine tumors in 2004 were removed in the 4<sup>th</sup> version published in 2017.<sup>[35, 36]</sup> Because all PCCs and PGLs may have metastatic potential. Therefore, in version 4, the "metastatic" term is used instead of "malignant" for both PCCs and PGLs. This also eliminated the confusion that may occur between locally invasive and distant metastatic tumors.<sup>[9]</sup> PGLs are divided into two groups as sympathetic paragangliomas and parasympathetic paragangliomas according to their clinical and biological behavior in version 4. Parasympathetic PGLs are often localized primarily in the head and neck region, rarely localized in the thorax and pelvis. For this reason, PGLs are examined under two subtitles as sympathetic PGLs and head and neck PGLs.<sup>[36]</sup>

#### Metastatic Pheochromocytomas/Paragangliomas

Although most of the PPGLs are benign, the metastatic disease may develop in 15-17%. Metastatic disease is reported between 2-25% in PCCs and 2.4-60% in PGLs.<sup>[37, 38]</sup>

Head and neck PGLs constitute 20% of all PGLs, but the rate of metastatic disease is 5%.<sup>[9]</sup> Synchronous metastasis is present in 35-52% of patients with metastatic PPGL and metachronous in 48-65%.<sup>[37, 38]</sup> While at least 30% of the patients with PPGL have a hereditary predisposition, almost 50% of metastatic patients have germline mutations.<sup>[39]</sup> In metastatic disease, male gender, advanced age, synchronous metastasis, larger primary tumor, patients without primary tumor resection, patients with sympathetic PGL and tumors with increased dopamine are associated with short-term survival.<sup>[37, 38]</sup> The mean tumor diameter is larger and 8.2 cm in patients with metastatic tumors. 16% of metastatic tumors are under 5 cm in size, most of

them (78.5%) are PGL, and only 21.5% of them are PCC.<sup>[37]</sup> Germline SDHB mutation is thought to be one of the most important risk factors for the development of metastases in patients with PPGL.<sup>[40]</sup> More than half of patients with metastatic PGL and 5-10% of patients with metastatic PCC have germline SDHB mutation. MAX mutation carriers have a 10-25% risk of developing metastatic disease, but MAX mutations are rare. VHL and SDHD mutation carriers have less than 5% risk of metastatic disease.[39] The course of metastatic disease is heterogeneous and the 5-year survival rate ranges between 40-85%.<sup>[33]</sup> The most common metastasis sites in metastatic PPGL are 80% local and distant lymph nodes, 72% bones, 50% liver, and 50% lungs.<sup>[39]</sup> Bone and lymph node metastasis in PGL is more than PCC, while liver metastasis is more common in PCC.<sup>[40]</sup> 91% of bone metastasis are lytic lesions, 81% are in vertebrae and 67% in the pelvis. Bone lesions were detected in CT or MRI with a rate of 81% and the detection rate in FDG-PET/CT is 95%. Survival in bone metastases is better than liver and lung metastases. Median total survival was 12 years in those with only bone metastasis, 7.5 years in liver and lung metastases, and five years in patients with bone, liver and lung metastases.<sup>[41]</sup> The TNM staging system of the American Joint Committee on Cancer (AJCC) was developed to predict the prognosis, based on the specific anatomical features of the primary tumor and the occurrence of metastasis. In the light of the data accumulated in the literature, for the first time, staging for PPGL was done in the updated version 8 of the TNM staging system (Table 2).<sup>[42]</sup> TNM staging system for PPGL was implemented on January 01, 2017. It will guide clinicians in determining the type and extension of surgery, indications for neoadjuvant and adjuvant treatments and the follow-up needed by metastatic PPGL patients. Comprehensive staging and survival data will be collected over the coming years. These data are expected to further our understanding of PPGL and facilitate the development of effective treatments.[40]

#### Disclosures

Peer-review: Externally peer-reviewed.

Conflict of Interest: None declared.

Authorship Contributions: Concept – M.U., N.A.; Design – M.U., N.A.; Supervision – M.U.; Materials – M.U., N.A.; Data collection &/or processing – M.U., N.A.; Analysis and/or interpretation – M.U.; Literature search – M.U., N.A.; Writing – M.U., N.A.; Critical review – M.U.

## References

- Tevosian SG, Ghayee HK. Pheochromocytomas and Paragangliomas. Endocrinol Metab Clin North Am 2019;48:727–50. [CrossRef]
- 2. Patel D, Phay JE, Yen TWF, Dickson PV, Wang TS, Garcia R, et al. Up-

Table 2. TNM classifications of pheochromocytoma and
paraganglioma

Primary Tumor (T)	
Tx	Tumor cannot be assessed
T1	<5 cm in greatest diameter, no
	extra-adrenal invasion
T2	>5 cm or sympathetic paraganglioma
	of any size, no extra-adrenal invasion
Т3	Any size with invasion into
	surrounding tissues (e.g., liver,
	pancreas, spleen, and and kidneys)
Nodal Involvement (N)	
Nx	Regional lymph nodes cannot be assessed
NO	No regional lymph node metastasis
N1	Regional lymph node metastasis
Distant metastases (M)	
MO	No distant metastasis
M1	Distant metastasis
M1a	Metastasis to only bone
M1b	Metastasis to only distant lymph
	nodes/liver or lung
M1c	Metastasis to the bone and multiple
	other sites
Stage Groups	
Stage 1	T1, N0, M0
Stage 2	T2, N0, M0
Stage 3	T1, N1, M0 or T2, N1, M0 or T3,
	Any N, M0
Stage 4	Any T, Any N, Any M1

date on Pheochromocytoma and Paraganglioma from the SSO Endocrine/Head and Neck Disease-Site Work Group. Part 1 of 2: Advances in Pathogenesis and Diagnosis of Pheochromocytoma and Paraganglioma. Ann Surg Oncol 2020;27:1329–37. [CrossRef]

- Schreiner F, Beuschlein F. Disease monitoring of patients with pheochromocytoma or paraganglioma by biomarkers and imaging studies. Best Pract Res Clin Endocrinol Metab 2019 Oct 21 [Epub ahead of print], doi: 10.1016/j.beem.2019.101347. [CrossRef]
- Lenders JW, Duh QY, Eisenhofer G, Gimenez-Roqueplo AP, Grebe SK, Murad MH, Naruse M, Pacak K, Young WF Jr; Endocrine Society. Pheochromocytoma and paraganglioma: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2014 Jun;99(6):1915-42. [CrossRef]
- Kiernan CM, Solórzano CC. Pheochromocytoma and Paraganglioma: Diagnosis, Genetics, and Treatment. Surg Oncol Clin N Am 2016;25:119–38. [CrossRef]
- 6. Chen H, Sippel RS, O'Dorisio MS, Vinik AI, Lloyd RV, Pacak K; North American Neuroendocrine Tumor Society (NANETS). The North American Neuroendocrine Tumor Society consensus guideline for the diagnosis and management of neuroendocrine tumors: pheochromocytoma, paraganglioma, and medullary thyroid can-

cer. Pancreas 2010;39:775-83. [CrossRef]

- Crona J, Taïeb D, Pacak K. New Perspectives on Pheochromocytoma and Paraganglioma: Toward a Molecular Classification. Endocr Rev 2017;38:489–515. [CrossRef]
- 8. Bholah R, Bunchman TE. Review of Pediatric Pheochromocytoma and Paraganglioma. Front Pediatr 2017;5:155. [CrossRef]
- Lam AK. Update on Adrenal Tumours in 2017 World Health Organization (WHO) of Endocrine Tumours. Endocr Pathol 2017;28:213–27. [CrossRef]
- Favier J, Amar L, Gimenez-Roqueplo AP. Paraganglioma and phaeochromocytoma: from genetics to personalized medicine. Nat Rev Endocrinol 2015;11:101–11. [CrossRef]
- 11. Graham JB. Pheochromocytoma and hypertension; an analysis of 207 cases. Int Abstr Surg 1951;92:105–21.
- Eisenhofer G, Huynh TT, Hiroi M, Pacak K. Understanding catecholamine metabolism as a guide to the biochemical diagnosis of pheochromocytoma. Rev Endocr Metab Disord 2001;2:297– 311. [CrossRef]
- Cisco RM, Grondal S, Hamberger B. Adrenal physiology. In: Clark OH, Duh QY, Kebebew E, Gosnell JE, Shen WT, editors. Textbook of Endocrine Surgery. 3rd ed. Jaypee Brothers Medical Publishers: 2016. p. 973–8. [CrossRef]
- 14. Lal G, Clark OH. Thyroid, parathyroid and adrenal. In: Brunucardi FC, Andersen DK, Billiar TR, Kao LS, Hunter JG, Matthews JB, et al, editors. Schwartz's Principles of Surgery. 11th ed. McGraw Hill: New York; 2019. p. 1625–704.
- Zuber SM, Kantorovich V, Pacak K. Hypertension in pheochromocytoma: characteristics and treatment. Endocrinol Metab Clin North Am 2011;40:295–311. [CrossRef]
- 16. Gunawardane PTK, Grossman A. Phaeochromocytoma and Paraganglioma. Adv Exp Med Biol 2017;956:239–59. [CrossRef]
- 17. Jochmanova I, Pacak K. Genomic Landscape of Pheochromocytoma and Paraganglioma. Trends Cancer 2018;4:6–9. [CrossRef]
- Khatami F, Mohammadamoli M, Tavangar SM. Genetic and epigenetic differences of benign and malignant pheochromocytomas and paragangliomas (PPGLs). Endocr Regul 2018;52:41–54.
- Fishbein L, Leshchiner I, Walter V, Danilova L, Robertson AG, Johnson AR, et al. Comprehensive Molecular Characterization of Pheochromocytoma and Paraganglioma. Cancer Cell 2017;31:181–93. [CrossRef]
- Nölting S, Ullrich M, Pietzsch J, Ziegler CG, Eisenhofer G, Grossman A, et al. Current Management of Pheochromocytoma/Paraganglioma: A Guide for the Practicing Clinician in the Era of Precision Medicine. Cancers (Basel) 2019;11:1505. [CrossRef]
- 21. Zhuang Z, Yang C, Lorenzo F, Merino M, Fojo T, Kebebew E, et al. Somatic HIF2A gain-of-function mutations in paraganglioma with polycythemia. N Engl J Med 2012;367:922–30. [CrossRef]
- Antonio K, Valdez MMN, Mercado-Asis L, Taïeb D, Pacak K. Pheochromocytoma/paraganglioma: recent updates in genetics, biochemistry, immunohistochemistry, metabolomics, imaging and therapeutic options. Gland Surg 2020;9:105–23. [CrossRef]

- 23. Raue F, Frank-Raue K. Update on Multiple Endocrine Neoplasia Type 2: Focus on Medullary Thyroid Carcinoma. J Endocr Soc 2018;2:933–43. [CrossRef]
- 24. Grogan RH. The Importance of Family History in the Management of Endocrine Disease. Surg Clin North Am 2019;99:711–20. [CrossRef]
- 25. lacobone M, Citton M, Viel G, Schiavone D, Torresan F. Surgical approaches in hereditary endocrine tumors. Updates Surg 2017;69:181–91. [CrossRef]
- Frunzac RW, Grant CS. Pheochromocytoma. In: Clark OH, Duh QY, Kebebew E, Gosnell JE, Shen WT, editors. Textbook of Endocrine Surgery. 3rd ed. Jaypee Brothers Medical Publishers: 2016. p. 1047–67. [CrossRef]
- 27. Asari R, Scheuba C, Kaczirek K, Niederle B. Estimated risk of pheochromocytoma recurrence after adrenal-sparing surgery in patients with multiple endocrine neoplasia type 2A. Arch Surg 2006;141:1199–205. [CrossRef]
- Boedeker CC, Erlic Z, Richard S, Kontny U, Gimenez-Roqueplo AP, Cascon A, et al. Head and neck paragangliomas in von Hippel-Lindau disease and multiple endocrine neoplasia type 2. J Clin Endocrinol Metab 2009;94:1938–44. [CrossRef]
- 29. Crespigio J, Berbel LCL, Dias MA, Berbel RF, Pereira SS, Pig¬natelli D, et al. Von Hippel-Lindau disease: A single gene, sev¬eral hereditary tumors. J Endocrinol Investig 2018;41:21–31. [CrossRef]
- 30. Shen MH, Harper PS, Upadhyaya M. Molecular genetics of neurofibromatosis type 1 (NF1). J Med Genet 1996;33:2–17. [CrossRef]
- Liu P, Li M, Guan X, Yu A, Xiao Q, Wang C, et al. Clinical Syndromes and Genetic Screening Strategies of Pheochromocytoma and Paraganglioma. J Kidney Cancer VHL 2018;5:14–22. [CrossRef]
- 32. Martucci VL, Pacak K. Pheochromocytoma and paraganglioma: diagnosis, genetics, management, and treatment. Curr Probl Cancer 2014;38:7–41. [CrossRef]
- Corssmit EPM, Snel M, Kapiteijn E. Malignant pheochromocytoma and paraganglioma: management options. Curr Opin Oncol 2020;32:20–6. [CrossRef]
- 34. Plouin PF, Amar L, Dekkers OM, Fassnacht M, Gimenez-Roqueplo AP, Lenders JW, et al; Guideline Working Group. European Society of Endocrinology Clinical Practice Guideline for long-term followup of patients operated on for a phaeochromocytoma or a paraganglioma. Eur J Endocrinol 2016;174:G1–10. [CrossRef]
- DeLellis RA, Lloyd RV, Heitz PU, Eng C. WHO classification of tumours: pathology and genetics of tumours of endocrine organs.
  3rd ed. Lyon: IARC; 2004.
- Lloyd RV, Osamura RY, Kloppel G, Rosai J. WHO classification of tumours: pathology and genetics of tumours of endocrine organs. 4th ed. Lyon: IARC; 2017.
- 37. Ayala-Ramirez M, Feng L, Johnson MM, Ejaz S, Habra MA, Rich T, et al. Clinical risk factors for malignancy and overall survival in patients with pheochromocytomas and sympathetic paragangliomas: primary tumor size and primary tumor location as prognostic indicators. J Clin Endocrinol Metab 2011;96:717–25.
- 38. Hamidi O, Young WF Jr, Iñiguez-Ariza NM, Kittah NE, Gruber L,

Bancos C, Tamhane S, Bancos I. Malignant Pheochromocytoma and Paraganglioma: 272 Patients Over 55 Years. J Clin Endocrinol Metab 2017;102:3296–305. [CrossRef]

- Jimenez C, Rohren E, Habra MA, Rich T, Jimenez P, Ayala-Ramirez M, et al. Current and future treatments for malignant pheochromocytoma and sympathetic paraganglioma. Curr Oncol Rep 2013;15:356–71. [CrossRef]
- Roman-Gonzalez A, Jimenez C. Malignant pheochromocytomaparaganglioma: pathogenesis, TNM staging, and current clinical trials. Curr Opin Endocrinol Diabetes Obes 2017;24:174–83.
- 41. Ayala-Ramirez M, Palmer JL, Hofmann MC, de la Cruz M, Moon BS, Waguespack SG, et al. Bone metastases and skeletal-related events in patients with malignant pheochromocytoma and sympathetic paraganglioma. J Clin Endocrinol Metab 2013;98:1492– 7. [CrossRef]
- 42. Jimenez C, Libutti SK, Landry CS, Lloyd RV, McKay RR, Rohren E, et al. Adrenal-neuroendocrine tumors. In: Amin MB, Edge S, Greene F, Byrd DR, Brookland RK, Washington MK, et al, editors. AJCC cancer staging manual. 8th ed. New York: Springer; 2017. p. 919–27.